



This is an open access article distributed in accordance with the Creative Commons Attribution (CC BY 4.0) license: <https://creativecommons.org/licenses/by/4.0/> which permits any use, Share — copy and redistribute the material in any medium or format, Adapt — remix, transform, and build upon the material for any purpose, as long as the authors and the original source are properly cited. © The Author(s) 2023

Trend of antimicrobial resistance among *Salmonella typhi* isolated from the pediatric population presenting at a tertiary care hospital in Lahore

Farhan Rasheed¹, Iqra Jamil^{2*}, Tahira Tehseen³, Ahmad Yar⁴, Farhana Ali⁵, Ameena Ashraf⁶

ABSTRACT

Background and Objective: Typhoid fever caused by *Salmonella typhi* (*S. typhi*) is an important cause of morbidity and mortality worldwide. The emergence of resistance against the first and second line of drugs has limited the range of drug choices for its treatment. Therefore, the present study was designed to assess the status of drug resistance in *S. typhi* isolated from the blood samples of pediatric patients reporting at a leading tertiary care hospital in Lahore.

Methods: This descriptive study was conducted at the Pathology Department of Allama Iqbal Medical College, Lahore, Pakistan, over a period of 2 years. A total of 1,306 blood samples were aseptically collected from the pediatric population of Jinnah Hospital, Lahore, Pakistan, after obtaining informed assent from the parents/guardians. The samples were transferred into a Bact/Alert blood culture bottle. After the detection of microbial growth by the Bact/Alert 3-D system, bacterial isolates were identified by standard microbiological procedures. VITEK 2 compact automated identification and antimicrobial susceptibility testing instrument (bioMerieux) was used for the antimicrobial sensitivity testing. Data were entered and analyzed by using Statistical Package for the Social Sciences version 21.0.

Results: A total of 235 out of 1,306 (17.9%) blood culture samples yielded bacterial growth, among which 62.5% ($n = 147$) were *S. typhi*. Among 147 *S. typhi* isolates, 70.6% were multidrug resistant, while 63% were extensively drug resistant (XDR). Maximum resistance against first-line drugs observed was for ampicillin (79.4) and co-trimoxazole (67.2).

Conclusion: The emergence and spread of XDR *S. typhi* with a high level of resistance are quite alarming, leaving limited treatment options for the pediatric population. Moreover, increasing resistance to antibiotics demonstrates the quick waning of the efficacy of available first and second-line drugs in the treatment of typhoid fever.

Keywords: *Salmonella typhi*, multidrug resistant (MDR), extensive-drug resistant (XDR), typhoid fever, antibiotics.

Received: 27 March 2023

Revised date: 01 May 2023

Accepted: 04 June 2023

Correspondence to: Iqra Jamil

*PhD Microbiology Scholar, Department of Industrial Biotechnology, Government College University, Lahore, Pakistan.

Email: iqrajameel@hotmail.com

Full list of author information is available at the end of the article.

Introduction

Typhoid fever caused by *Salmonella enterica* serovar *typhi* is a systemic infection resulting in an increased morbidity and mortality rate throughout the world. About 10.9 million new cases and 116,800 typhoid fever-related deaths are reported to be occurring per year globally.^{1,2} According to the World Health Organization (WHO), typhoid is a serious public health issue that primarily affects children and young people.³ The third-highest typhoid rate in the world is reported in Pakistan. In 2020, the prevalence rate of typhoid fever affecting children between the ages of 2 and 5 years was 573.2 per 100,000 population per year in Pakistan.⁴ Route of transmission is mainly oral-fecal for enteric fever;

therefore, the consumption of fecal-contaminated water and food plays a major role in its spread.⁵

Ampicillin, co-trimoxazole, and chloramphenicol were considered to be the first-line antibiotics for typhoid fever treatment,⁶ but during the last few decades, the emergence of multidrug resistance (MDR) in *S. typhi* strains, particularly in developing countries, has been reported in escalation. MDR *S. typhi* strains are generally regarded to be resistant against at least one of the three or more than three categorically differentiated antimicrobials, such as ampicillin, sulfonamides (trimethoprim-sulfamethoxazole), and chloramphenicol, whereas extensively drug-resistant (XDR)

are those which are observed to be resistant to all but one or two antimicrobials, demonstrating resistance against several types of antibiotics, such as chloramphenicol, ampicillin, sulfonamides, fluoroquinolones (ciprofloxacin), and third-generation cephalosporins (ceftazidime, cefuroxime, and ceftriaxone), leaving out few options, such as piperacillin/tazobactam, azithromycin, and carbapenems, as efficient treatment options.^{7,8} Acquisition of resistant genes in *S. typhi* strains is mainly caused by transmissible plasmids. The first isolate of MDR *S. typhi* was reported between 1970 and 1980, which exhibited resistance to first-line drugs (ampicillin, co-trimoxazole, and chloramphenicol).⁹ Thereafter, ciprofloxacin became the preferred drug for *S. typhi* infections, but resistance developed quickly. The first case of ciprofloxacin resistance was reported in 1991.¹⁰ However, the incidence of MDR-*S. typhi* increased from 34.2% to 48.5% during the period from 2001 to 2006 as reported by the Aga Khan University in Pakistan. During this period, ciprofloxacin resistance also raised from 1.6% to 64.1%.¹¹

Third-generation cephalosporins, macrolides, and carbapenems have been used more frequently to treat typhoid fever since the development of MDR *S. typhi* and fluoroquinolone resistance. In Pakistan, the first case of XDR *S. typhi* was reported in Hyderabad in 2016. XDR strain showed resistance against first-line drugs, a fluoroquinolone, and a third-generation cephalosporin.^{12,13} Since then, more than 10,365 infections with XDR *S. typhi* have been reported from Pakistan, as per WHO reports.¹⁴ Despite the efforts made by infection control programs to control the disease, there was a gradual increase in documented cases over a 2-year period (2017 to 2018).¹⁴ Therefore, the present study was designed to determine the frequency and current status of drug resistance of *S. typhi* isolated from the blood samples of pediatric patients with septicemia presenting at one of the largest tertiary care hospitals in Lahore, Pakistan.

Methods

The present study was conducted at the Pathology Department of Allama Iqbal Medical College, Lahore, Pakistan, over a period of 2 years, from January 1, 2021 to December 31, 2022. Samples were collected from the college associated with Jinnah Hospital, Lahore. Samples from both genders of pediatric patients, suspected of bacterial sepsis and presenting with fever,¹⁵ within the age group of 0-12 years, were included in the study. Repetitive samples from the same patient were excluded. Also, patients with bleeding disorders, multiple co-morbidities, having any malignancy, or who were already on antibiotic therapy were excluded. A total of 1,306 blood cultures from suspected cases of bacterial sepsis were analyzed. Blood samples (4 ml) were collected in Bact/Alert blood culture bottles from each patient by an

experienced staff nurse. The samples were transported to the Microbiology Laboratory. Blood culturing was done through automation in the Bact/Alert 3-D culture system (bioMerieux, Durham, NC).¹⁶ After the detection of microbial growth, Gram staining was performed on the positive samples. Samples were sub-cultured on blood agar (Oxoid Ltd., Basingstoke, UK), chocolate agar (Oxoid Ltd., Basingstoke, UK), and MacConkey agar (Oxoid Ltd., Basingstoke, UK). After sub-culturing, the inoculated culture plates were incubated for 18-24 hours at 37°C aerobically. After overnight incubation, different microbiological procedures were used to identify bacterial isolates.¹⁷ For the identification of Gram-negative bacteria, Analytical Profile Index 20E was used.¹⁸ VITEK 2 compact automated identification and antimicrobial susceptibility testing instruments (bioMerieux) were used for the antimicrobial sensitivity testing.¹⁹ The isolates were labeled “multidrug-resistant” (MDR), which was resistant to first line drugs, and “extensively drug-resistant” (XDR), which showed resistance to more than two classes of antibiotics.²⁰

Statistical analysis

Data were entered and analyzed by using Statistical Package for the Social Sciences version 21.0. Sensitivity patterns were presented as frequency and percentages. Cross tabulation was done for gender and age. The chi-square test was used to assess statistical significance with a *p*-value < 0.05 taken as statistically significant.

Results

Out of 1,306 blood cultures, 235 (17.9%) were positive for bacterial growth. Out of these 235 positive samples, 62.5% yielded *S. enterica* species (72.1% were *S. typhi* while 27.9% were *S. paratyphi A*). The sensitivity pattern of all the isolates (*n* = 147) indicated 70.6% were MDR while 63% were XDR *S. enterica* isolates (Figure 1).

Antimicrobial susceptibility testing revealed that 79.4%, 74.7%, and 67% of *S. typhi* were resistant to ampicillin, ciprofloxacin, and co-trimoxazole, respectively, while 65.4% isolates were resistant to chloramphenicol. All isolates of *S. paratyphi A* tested for azithromycin were susceptible (Table 1).

Discussion

Enteric fever is a serious public health issue across the world, putting a significant burden on healthcare system, especially in developing countries. In Pakistan, an alarming situation regarding antibiotic resistance has been developed because of the increase in the emergence of MDR and XDR strains.⁵ In the present study, 72.1% (*n* = 106) isolates were of *S. typhi* out of 147 *S. enterica* species. Similar results were reported by Khanal et al.²¹ from B. P. Koirala Institute of Health Sciences Hospital, Nepal, in which *S. typhi* was the most

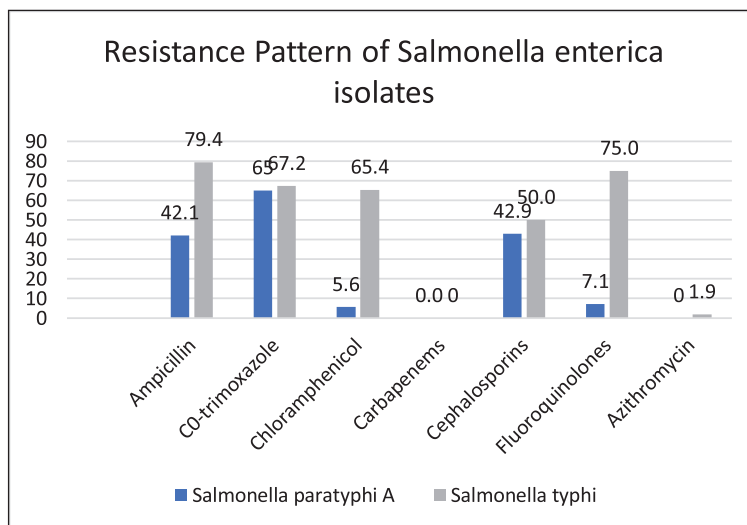


Figure 1. Resistance pattern of *S. enterica* serovars against different classes of antimicrobial drugs. Resistance of *S. typhi* against ampicillin was 79.4%, while 75% and 67.2% of isolates were resistant to fluoroquinolones and co-trimoxazole, respectively. 65% *S. paratyphi A* isolates were resistant to co-trimoxazole. Carbapenems showed 100% sensitivity against *S. typhi* and *S. paratyphi A*.

Table 1. Antimicrobial sensitivity pattern of *S. typhi* among 147 isolates.

Antibiotics	Sensitive (%)	Resistant (%)
First line drugs		
1. Ampicillin	22 (20.6)	85 (79.4) ^a
2. CO-trimoxazole	35 (32.8)	72 (67.2)
3. Chloramphenicol	37 (34.6)	70 (65.4)
Second line drugs		
1. Ciprofloxacin	27 (25.3)	80 (74.7)
2. Ceftriaxone	54 (46.8)	53 (53.2)
3. Cefipime	62 (58)	45 (42)
Third line drugs ^a		
1. Imipenem	107 (100)	0 (0)
2. Meropenem	107 (100)	0 (0)

^aMaximum resistance was observed against ampicillin (79.4%), while 100% isolates were sensitive against third line of drugs.

common serotype isolated from the blood samples from the patients with enteric fever. Another study by Zakir et al.²² from Pakistan in 2021 also reported *S. typhi* as the most common serotype (90%) isolated from patients with enteric fever. In accordance with our study, Fatima et al.²³ and Qamar et al.²⁴ from Pakistan also reported similar results.

Various studies from all over Pakistan have documented the presence of MDR *S. typhi*.²¹⁻²³ MDR *S. typhi* are found to be resistant against first-line antibiotics, which include ampicillin, chloramphenicol, and trimethoprim-sulfamethoxazole. In the present study, 70.6% of isolates were found to be MDR. A retrospective study conducted by Klemm et al.,¹² conducted over a period of 9 years, also reported a high burden of MDR and fluoroquinolones-resistant strains. Blood samples

were collected from tertiary care hospitals of Karachi and Hyderabad from the Sindh region of Pakistan. Similar results were also reported by Park et al.,²⁵ who showed a rapid increase in MDR *S. typhi* in Ghana, Kenya, and Tanzania, and reported 52% MDR *S. typhi* isolates among the total number of *S. typhi* strains across the samples collected from 11 countries of Sub Saharan Africa. Misuse or uncontrolled use of antimicrobials that are sold over the counter without a prescription or medical supervision may be the root cause of the emergence of MDR strains.

In Pakistan, the majority of MDR strains of *S. typhi* have attained additional resistance to fluoroquinolones and beta-lactams, which marked them as XDR.¹² In the current study, 63% isolates were found to be XDR, which were in accordance with another study conducted in Pakistan which reported 54% XDR strains.²⁶ Similar to our results, many local and international studies also reported a high prevalence of XDR strains.²⁵⁻²⁹ In the current study, 79.4% and 67% of *S. typhi* were resistant to ampicillin and co-trimoxazole, respectively. A study conducted in China documented ascending trends in resistance to ampicillin (91%) and co-trimoxazole (88%) in *S. typhi* isolated from blood samples from patients with enteric fever.³⁰ The two drugs ceftriaxone and ciprofloxacin are regarded as the main pillars of typhoid treatment, especially in developing countries, but in the present study, high resistance was also observed against these antibiotics. A total of 75% of isolates showed resistance against ciprofloxacin in the current study, which is in accordance with other similar studies published locally and internationally.^{12,29,31,32} Similar results were also reported in a study conducted in Taiwan,

Table 2. Antimicrobial sensitivity pattern of *S. typhi* with respect to gender (cross tabulation).

Category	Drugs	Male		Female		p-value*
		Sensitive (%)	Resistant (%)	Sensitive (%)	Resistant (%)	
First line drugs	Ampicillin	19.8%	80.2%	21.5%	78.5%	0.08
	Co-trimoxazole	36.6%	63.4%	29%	71%	
	Chloramphenicol	35.8%	64.2%	33.4%	66.6%	
Second line drugs	Ciprofloxacin	24.3%	75.7%	26.3%	73.7%	0.06
	Ceftriaxone	45.8%	54.2%	47.8%	52.2%	
	Cefipime	54.5%	45.5%	61.5%	38.5%	
Third line drugs	Imipenem	100%	0%	100%	0%	0.1
	Meropenem	100%	0%	100%	0%	

*Statistically insignificant results reflect no association between gender and resistance pattern of *S. typhi*.

Table 3. Antimicrobial sensitivity pattern of *S. typhi* with respect to age (cross tabulation).

Category	Drugs	<10 years		>10 years		p-value*
		Sensitive (%)	Resistant (%)	Sensitive (%)	Resistant (%)	
First line drugs	Ampicillin	18.8	81.2	22.4	77.6	0.07
	Co-trimoxazole	34.6	65.4	31	69	
	Chloramphenicol	36.8	63.2	32.4	67.6	
Second line drugs	Ciprofloxacin	24.8	75.2	25.9	74.1	0.06
	Ceftriaxone	44.8	55.2	48.8	51.2	
	Cefipime	53.8	46.2	62.2	37.8	
Third line drugs	Imipenem	100	0.0	100	0.0	0.12
	Meropenem	100	0.0	100	0.0	

*Statistically insignificant results depict no association between age and resistance pattern of *S. typhi*.

which showed 69% resistance against ciprofloxacin.³³ Association of the resistance pattern of *S. typhi* with gender and age was found to be statistically insignificant in the present study (Tables 2 and 3), which depicts that gender and age are independent of the antimicrobial resistance pattern in the study population.

Regarding third-line drugs for enteric fever treatment, carbapenems showed 100% sensitivity against *S. enterica* serovars. Azithromycin was also highly sensitive against *S. enterica* isolates. A study conducted by Sjölund-Karlsson et al.³⁴ also reported 93.6% sensitivity to azithromycin against *S. typhi* isolated from the blood samples of enteric fever patients. Nowadays, azithromycin is the drug of choice for treating uncomplicated enteric fever. The prospect of antibiotic resistance, however, necessitates continuous monitoring of its sensitivity patterns.

While focusing on increasing antimicrobial resistance, actions must be taken to raise health and hygiene awareness among health practitioners as well as for the general population. In developing countries such as Pakistan, where antibiotic stewardship programs are not up to date, this should be a serious concern for organizations working on it.

Various practices, including good hand hygiene, improved sanitation, and effective antibiotic policy, can help to control the disease and thereby reduce antimicrobial resistance in the community. To reduce the burden of disease caused by *S. typhi*, effective surveillance, improved diagnostics, more rational antimicrobial usage, and effective mass vaccination will all be crucial.

Conclusion

The emergence and spread of XDR *S. typhi* with a high level of resistance are quite alarming, leaving limited treatment options for the pediatric population in Pakistan. Moreover, increasing resistance to antibiotics demonstrates the quick waning of the efficacy of available first- and second-line drugs in the treatment of typhoid fever. Antibiotic stewardship, continuous surveillance, and vaccination on a mass level should be the priority areas to reduce the spread of this deadly disease.

Limitations of the Study

This was a single-center study; because of the extensive spread of XDR *S. typhi*, a multicenter study should be conducted to determine

a more exploratory and representative picture of the resistance pattern in our region.

Acknowledgement

The authors would like to acknowledge all the respondents who participated in the study, including the staff of the Pathology department of Allama Iqbal Medical College Lahore, Pakistan, who processed the samples.

List of Abbreviations

ID/AST	Identification and antimicrobial susceptibility testing
MDR	Multidrug resistant
<i>S. typhi</i>	<i>Salmonella typhi</i>
WHO	World Health Organization
XDR	Extensively drug resistant

Conflict of interest

None to declare.

Grant support and financial disclosure

None to disclose.

Ethical approval

The study was approved by the Ethical Review Board of Allama Iqbal Medical College/Jinnah Hospital, Lahore, Pakistan, vide reference no: 47/ERB/2019, dated March 13th, 2019.

Authors' contributions

FR: Concept and design of study, analysis of data, critical intellectual input to the manuscript.

IJ, TT, AY: Drafting the manuscript, acquisition of data.

FA, AA: critical intellectual input to the manuscript.

ALL AUTHORS: Approval of the final version of the manuscript to be published.

Authors' Details

Farhan Rasheed¹, Iqra Jamil², Tahira Tehseen³, Ahmad Yar⁴, Farhana Ali⁵, Ameena Ashraf⁶

1. Associate Professor, Department of Pathology, Allama Iqbal Medical College, Lahore, Pakistan
2. Ph.D Microbiology Scholar, Department of Industrial Biotechnology, Government College University, Lahore, Pakistan
3. Assistant Professor, Department of Pathology, Wah Medical College, Wah Cantt, Pakistan
4. Classified Pathologist, Combined Military Hospital, Sialkot, Pakistan
5. Assistant Professor, Department of Pathology, University of Child Health Sciences, The Children's Hospital, Lahore, Pakistan
6. Professor, Department of Pathology, Allama Iqbal Medical College, Lahore, Pakistan

References

1. Stanaway JD, Reiner RC, Blacker BF, Goldberg EM, Khalil IA, Troeger CE, et al. The global burden of typhoid and paratyphoid fevers: a systematic analysis for the Global Burden of Disease Study 2017. *Lancet Infect Dis*. 2019 Apr 1;19(4):369–81. [https://doi.org/10.1016/S1473-3099\(18\)30685-6](https://doi.org/10.1016/S1473-3099(18)30685-6).
2. Marchello CS, Hong CY, Crump JA. Global typhoid fever incidence: a systematic review and meta-analysis. *Clin Infect Dis*. 2019 Mar 7;68(Supplement_2):S105–16. <https://doi.org/10.1093/cid/ciy1094>
3. Steele AD, Hay Burgess DC, Diaz Z, Carey ME, Zaidi AK. Challenges and opportunities for typhoid fever control: a call for coordinated action. *Clin Infect Dis*. 2016 Mar 15;62(suppl_1):S4–8. <https://doi.org/10.1093/cid/civ976>
4. Akram J, Khan AS, Khan HA, Gilani SA, Akram SJ, Ahmad FJ, et al. Extensively drug-resistant (XDR) typhoid: evolution, prevention, and its management. *BioMed Res Int*. 2020 May 2;2020:6432580. <https://doi.org/10.1093/cid/civ976>
5. Radhakrishnan A, Als D, Mintz ED, Crump JA, Stanaway J, Breiman RF, et al. Introductory article on global burden and epidemiology of typhoid fever. *Am J Trop Med Hyg*. 2018 Sep;99(3 Suppl):4. <https://doi.org/10.4269/ajtmh.18-0032>
6. Crump JA, Sjölund-Karlsson M, Gordon MA, Parry CM. Epidemiology, clinical presentation, laboratory diagnosis, antimicrobial resistance, and antimicrobial management of invasive *Salmonella* infections. *Clin Microbiol Rev*. 2015 Oct;28(4):901–37. <https://doi.org/10.1128/cmr.00002-15>
7. Basak S, Singh P, Rajurkar M. Multidrug resistant and extensively drug resistant bacteria: a study. *J Pathog*. 2016 Oct;2016:4065603. <https://doi.org/10.1155/2016/4065603>
8. Saeed M, Rasool MH, Rasheed F, Saqalein M, Nisar MA, Imran AA, et al. Extended-spectrum beta-lactamases producing extensively drug-resistant *Salmonella typhi* in Punjab, Pakistan. *J Infect Dev Ctries*. 2020 Feb 29;14(02):169–76. <https://doi.org/10.3855/jidc.12049>
9. Dyson ZA, Klemm EJ, Palmer S, Dougan G. Antibiotic resistance and typhoid. *Clin Infect Dis*. 2019 Mar 7;68(Supplement_2):S165–70. <https://doi.org/10.1093/cid/ciy1111>
10. Umasankar S, Wall RA, Berger J. A case of ciprofloxacin-resistant typhoid fever. *Commun Dis Rep CDR Rev*. 1992 Nov 1;2(12):R139–40.
11. Hasan R, Zafar A, Abbas Z, Mahraj V, Malik F, Zaidi A. Antibiotic resistance among *Salmonella enterica* serovars *typhi* and *paratyphi A* in Pakistan (2001-2006). *J Infect Dev Ctries*. 2008 Aug 1;2(04):289–94. <https://doi.org/10.3855/jidc.224>
12. Klemm EJ, Shakoore S, Page AJ, Qamar FN, Judge K, Saeed DK, et al. Emergence of an extensively drug-resistant *Salmonella enterica* serovar *typhi* clone harboring a promiscuous plasmid encoding resistance to fluoroquinolones and third-generation cephalosporins. *mBio*. 2018 Mar 7;9(1):e00105–18. <https://doi.org/10.1128/mbio.00105-18>
13. Qamar FN, Yousafzai MT, Khalid M, Kazi AM, Lohana H, Karim S, et al. Outbreak investigation of ceftriaxone-resistant *Salmonella enterica* serotype *typhi* and its risk factors among the general population in Hyderabad, Pakistan: a matched case-control study. *Lancet Infect Dis*. 2018 Dec 1;18(12):1368–76. [https://doi.org/10.1016/S1473-3099\(18\)30483-3](https://doi.org/10.1016/S1473-3099(18)30483-3)
14. Majeed MM, Munir A. Pakistan: country report on children's environmental health. *Rev. Environ Health*. 2020 Mar 26;35(1):57–63. <https://doi.org/10.1515/reveh-2019-0087>
15. Al-Mulla NA, Taj-Aldeen SJ, El Shafie S, Janahi M, Al-Nasser AA, Chandra P. Bacterial bloodstream infections and antimicrobial susceptibility pattern in pediatric hematology/oncology patients after anticancer chemotherapy. *Infect Drug Resist*. 2014 Nov 6:289–99.
16. Butt T, Afzal RK, Ahmad RN, Salman M, Mahmood A, Anwar M. Bloodstream infections in febrile neutropenic patients: bacterial spectrum and antimicrobial susceptibility pattern. *J Ayub Med Coll Abbottabad*. 2004;16(1):18–22.

17. Kara Ö, Zarakolu P, Aşçıoğlu S, Etgül S, Uz B, Büyükaşık Y, et al. Epidemiology and emerging resistance in bacterial bloodstream infections in patients with hematologic malignancies. *Infect Dis Rev*. 2015 Oct 3;47(10):686–93. <https://doi.org/10.3109/23744235.2015.1051105>
18. Wasihun AG, Wlekidan LN, Gebremariam SA, Dejene TA, Welderufael AL, Haile TD, et al. Bacteriological profile and antimicrobial susceptibility patterns of blood culture isolates among febrile patients in Mekelle Hospital, Northern Ethiopia. *Springerplus*. 2015 Dec;4(1):1–7. <https://doi.org/10.1186/s40064-015-1056-x>
19. Gupta S, Kashyap B. Bacteriological profile and antibiogram of blood culture isolates from a tertiary care hospital of North India. *Trop J Med Res*. 2016 Jul 1;19(2):94–9.
20. Sharma P, Dahiya S, Manral N, Kumari B, Kumar S, Pandey S, et al. Changing trends of culture-positive typhoid fever and antimicrobial susceptibility in a tertiary care North Indian Hospital over the last decade. *Indian J Med Microbiol*. 2018 Jan 1;36(1):70–6. https://doi.org/10.4103/ijmm.IJMM_17_412
21. Khanal B, Sharma SK, Bhattacharya SK, Bhattarai NR, Deb M, Kanungo R. Antimicrobial susceptibility patterns of *Salmonella enterica* serotype *typhi* in eastern Nepal. *J Health Popul Nutr*. 2007 Mar;25(1):82.
22. Zakir M, Khan M, Umar MI, Murtaza G, Ashraf M, Shamim S. Emerging trends of multidrug-resistant (MDR) and extensively drug-resistant (XDR) *Salmonella typhi* in a tertiary care Hospital of Lahore, Pakistan. *Microorganisms*. 2021 Nov 30;9(12):2484. <https://doi.org/10.3390/microorganisms9122484>
23. Fatima G, Kazmi SS, Kainat S. XDR/MDR *Salmonella*: an experience from a tertiary care hospital, Karachi, Pakistan. *Int J Infect Dis*. 2020 Dec 1;101:37. <https://doi.org/10.1016/j.ijid.2020.09.131>
24. Qamar FN, Yousafzai MT, Dehraj IF, Shakoor S, Irfan S, Hotwani A, et al. Antimicrobial resistance in typhoidal *Salmonella*: surveillance for enteric fever in Asia project, 2016–2019. *Clin Infect Dis*. 2020 Nov 1;71(Supplement_3):S276–84. <https://doi.org/10.1093/cid/ciaa1323>
25. Park SE, Pham DT, Boinett C, Wong VK, Pak GD, Panzner U, et al. The phylogeography and incidence of multi-drug resistant typhoid fever in sub-Saharan Africa. *Nat Commun* 2018 Nov 30;9(1):5094. <https://doi.org/10.1038/s41467-018-07370-z>
26. Batool A, Yunus N, Yaqoob A, Lone D, Khalid A, Ejaz H, et al. Prevalence of multi-drug resistant and extensively drug-resistant *Salmonella enterica* serovar *typhi* recovered from pediatrics' septicemia patients in Lahore. *Pak J Med Health Sci*. 2021;15(4):843–5.
27. Ejaz A, Khawaja A, Fatima K, Alavi N, Asif M. Frequency and antimicrobial resistance patterns of *Salmonella enterica* isolates in a tertiary care setting. *Pak J Health Sci*. 2022 May 26;16(05):11–3. <https://doi.org/10.53350/pjmhs2216511>
28. Jain S, Chugh TD. Antimicrobial resistance among blood culture isolates of *Salmonella enterica* in New Delhi. *J Infect Dev Ctries*. 2013 Nov 15;7(11):788–95. <https://doi.org/10.3855/jidc.3030>
29. Ramachandran A, Shanthi M, Sekar U. Detection of blaCTX-M extended spectrum beta-lactamase producing *Salmonella enterica* serotype *typhi* in a tertiary care centre. *J Clin Diagn Res*. *JCDR*. 2017 Sep;11(9):DC21. <https://doi.org/10.7860/JCDR/2017/30150.10637>
30. Chen HM, Wang Y, Su LH, Chiu CH. Nontyphoid *Salmonella* infection: microbiology, clinical features, and antimicrobial therapy. *Pediatr Neonatol*. 2013 Jun 1;54(3):147–52. <https://doi.org/10.1016/j.pedneo.2013.01.010>
31. Ali A, Ali HA, Shah FH, Zahid A, Aslam H, Javed B. Pattern of antimicrobial drug resistance of *Salmonella typhi* and *paratyphi A* in a Teaching Hospital in Islamabad. *J Pak Med Assoc*. 2017 Mar 1;67(3):375–9.
32. Qamar FN, Azmatullah A, Kazi AM, Khan E, Zaidi AK. A three-year review of antimicrobial resistance of *Salmonella enterica* serovars *typhi* and *paratyphi A* in Pakistan. *J Infect Dev Ctries*. 2014 Aug 13;8(08):981–6. <https://doi.org/10.3855/jidc.3817>
33. Lee CJ, Su LH, Huang YC, Chiu CH. First isolation of ciprofloxacin-resistant *Salmonella enterica* serovar *typhi* in Taiwan. *J Microbiol Immunol Infect*. 2013 Dec 1;46(6):469–73. <https://doi.org/10.1016/j.jmii.2013.01.002>
34. Sjölund-Karlsson M, Joyce K, Blickenstaff K, Ball T, Haro J, Medalla FM, et al. Antimicrobial susceptibility to azithromycin among *Salmonella enterica* isolates from the United States. *Antimicrob Agents Chemother*. 2011 Sep;55(9):3985–9. <https://doi.org/10.1128/aac.00590-11>